Article

A New Route to Diverse 1-Azasugars from N-Boc-5-hydroxy-3-piperidene as a Common Building Block

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A new general method for the synthesis of a variety of 1-azasugars with a nitrogen atom at the anomeric position is described. The readily available chiral N-Boc-5-hydroxy-3-piperidene 3 is transformed to isofagomine (2), homoisofagomine (13), and 5'-deoxyisofagomine (14) via stereoselective epoxidation and regioselective ring-cleavage in a highly stereocontrolled manner. In addition, the synthesis of all four stereoisomers of 3.4.5-trihydroxypiperidines (18-21) classified as 1-azasugar-type glycosidase inhibitors was stereoselectively achieved from the (chiral) piperidene **3**.

Introduction

Glycosciences are emerging as a key research field at the frontiers of biology, synthetic and supramolecular chemistry, and enzymology.¹ Among the carbohydrate processing enzymes, glycosidases have been identified as an important class of therapeutic targets with applications in the treatment of influenza infection,² cancer,³ AIDS,⁴ and diabetes.⁵ As a result, numerous classes of inhibitors have been developed, some of which provide interesting insights into the mechanism of enzymatic glycoside hydrolysis. Among them, both naturally occur-

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ring and synthetic polyhydroxylated piperidines⁶ have been shown to be specific and potent inhibitors of glycosidases⁷ and have been demonstrated to have great potential as drugs for treating a variety of carbohydratemediated diseases.⁸

The transition states for enzymatic glycosidase hydrolysis have considerable oxocarbenium-ion characteristics, in that the anomeric carbon acquires sp² hybridization and a partial positive charge develops at the anomeric carbon and the endocyclic oxygen.⁹ Iminosugars such as 1-deoxynojirimycin (1) and isofagomine (2), as transition-state analogues, are of particular interest in terms of inhibitor design (Figure 1). A new class of sugar mimic inhibitor having a nitrogen atom at the anomeric position is 1-azasugars such as 2. These compounds are

^{(1) (}a) Sears, P.; Wong, C.-H. Science 2001, 291, 2344-2350. (b)

Bertozzi, C. R.; Kiessling, L. *Science* **2001**, *291*, 2357–2364. (2) (a) von Itzstein, M.; Wu, W.-Y.; Kok, G. B.; Pegg, M. S.; Dyason, J. C.; Jin, B.; Phan, T. V.; Smythe, M. L.; White, H. F.; Oliver, S. W.; Colman, P. M.; Varghese, J. N.; Ryan, D. M.; Woods, J. M.; Bethell, R. C.; Hothman, V. J.; Cameron, J. M.; Penn, C. R. *Nature* **1993**, *363*, 418–423. (b) Kim, C. U.; Lew, W.; Williams, M. A.; Liu, H.; Zhang, L.; Swaminathan, S.; Bischofberger, N.; Chen, M. S.; Mendel, D. B.; Tai, C. Y.; Laver, W. G.; Stevens, R. C. J. Am. Chem. Soc. **1997**, *119*, 681– 690. (c) Chand, P.; Kotian, P. L.; Dehghani, A.; El-Kattan, Y.; Lin, T.-H.; Hutchison, T. L.; Sudhakar Babu, Y.; Bantia, S.; Elliott, A. J.; Montgomery, J. A. J. Med. Chem. 2001, 44, 4379-4392.

^{(3) (}a) Goss, P. E.; Baker, M. A.; Carver, J. P.; Dennis, J. W. Clin. Cancer Res. 1995, 1, 935-944. (b) Nishimura, Y.; Satoh, T.; Adachi, H.; Kondo, S.; Takeuchi, T.; Azetaka, M.; Fukuyasu, H.; Iizuka, Y. J. Med. Chem. **1997**, 40, 2626–2633. (c) Zitzmann, N.; Mehta, A. S.; Carrouée, S.; Butters, T. D.; Platt, F. M.; McCauley, J.; Blumberg, B. S.; Dwek, R. A.; Block, T. M. Proc. Natl. Acad. Sci. U.S.A. 1999, 96, 11878-11882.

^{(4) (}a) Karlsson, G. B.; Butters, T. D.; Dwek, R. A.; Platt, F. M. J. Biol. Chem. **1993**, 268, 570–576. (b) Groopman, J. E. Rev. Infect. Dis. 1990, 12, 931-937.

^{(5) (}a) Treadway, J. L.; Mendys, P.; Hoover, D. J. Expert Opin. Invest. Drugs 2001, 10, 439–454. (b) Jacob, G. S. Curr. Opin. Struct. Biol. 1995, 5, 605–611.

^{(6) (}a) Asano, N. Glycobiology 2003, 13, 93R-104R. (b) Asano, N. Curr. Top. Med. Chem. 2003, 3, 471-484. (c) Compain, P.; Martin, O. R. Curr. Top. Med. Chem. 2003, 3, 541-560. (d) Nash, R. J.; Watson, A. A.; Asano, N. In Alkaloids: Chemical and Biological Perspectives; Pelletier, S. W., Ed.; Elsevier: Oxford, UK, 1996; Vol. 11, pp 345-776. (e) Elbein, A. D.; Molyneux, R. J. In *Comprehensive Natural Products*; Barton, D., Nakanishi, K., Eds.; Elsevier: New York, 1999; Vol. 3, pp 129–160.

^{(7) (}a) Bols, M. Acc. Chem. Res. 1998, 31, 1. (b) Iminosugars as Glycosidase Inhibitors. Nojirimicin and Beyond; Stütz, A. E., Ed.;

<sup>Giycostalase Innibitors. Notritutti and Beyond; Statz, A. E., Ed.;
Wiley-VCH: Weinheim, Germany, 1999.
(8) (a) Somsak, L.; Nagy, V.; Hadady, Z.; Docsa, T.; Gergely, P. Curr.
Pharm. Des. 2003, 9, 1177–1189. (b) Greimel, P.; Spreitz, J.; Statz,
A. E.; Wrodnigg, T. M. Curr. Top. Med. Chem. 2003, 3, 513–523. (c)
Nishimura, Y. Curr. Top. Med. Chem. 2003, 3, 575–591.</sup>

 ^{(9) (}a) Zechel, D. L.; Boraston, A. B.; Gloster, T.; Boraston, C. M.;
 MacDonald, J.; Tilbrook, D. M. G.; Stick, R. V.; Davies, G. J. J. Am. Chem. Soc. 2003, 125, 14313-14323. (b) Zechel, D. L.; Withers, S. G. Acc. Chem. Res. 2000, 33, 11-18.



FIGURE 1. Transition state for the hydrolysis step of an inverting glycosidase and azasugars.

highly potent inhibitors of β -glycosidases, whose substrates they mimic.¹⁰

As a consequence, in recent years there has been a great deal of interest not only in the synthesis of the natural products themselves but also in the nature of chemically modified analogues. However, most of the methodologies described for the synthesis of these compounds, which can be regarded as azasugars (also called iminosugars), start from carbohydrates and, in general, require a large number of steps to reach a specific target. Thus, the development of new methods for the enantio-selective synthesis of polyhydroxylated piperidines constitutes an area of considerable current interest.¹¹

As a part of our ongoing interest in the polyhydroxy piperidines,¹² we envisioned the use of *N*-Boc-5-hydroxy-3-piperidene (**3**) as a general representative chiral building block that might permit easy access to these classes of compounds. This design was surmised keeping in mind the general substitution pattern of the 1-azasugars of **4**.

(11) (a) Kazmaier, U.; Schneider, C. Tetrahedron Lett. 1998, 39, 817–818. (b) Kim, Y. J.; Ichikawa M.; Ichikawa, Y. J. Org. Chem. 2000, 65, 2599–2602. (c) Mehta, G.; Mohal, N. Tetrahedron Lett. 2000, 41, 5747–5751. (d) Liang, X.; Lohse, A.; Bols, M. J. Org. Chem. 2000, 65, 7432–7437. (e) Pandey, G.; Kapur, M. Tetrahedron Lett. 2000, 41, 8821–8824. (f) Zhao, G.; Deo, U. C.; Ganem, B. Org. Lett. 2001, 3, 201–203. (g) Pandey, G.; Kapur, M. Synthesis 2001, 1263–1267. (h) Andersch, J.; Bols, M. Chem. Eur. J. 2001, 7, 3744–3747. (i) Comins, D. L.; Fulp, A. B. Tetrahedron Lett. 2001, 42, 6839–6841. (j) Best, W. M.; MacDonald, J. M.; Skelton, B. W.; Stick, R. V.; Tilbrook, D. M. G.; White, A. H. Can. J. Chem. 2002, 80, 857–865. (k) Guanti, G.; Riay, R. Tetrahedron Lett. 2003, 44, 357–360. (k) Pandey, G.; Kapur, M.; Khan, M. I.; Gaikwad, S. M. Org. Biomol. Chem. 2003, 1, 3321–3326.

(12) (a) Takahata, H.; Banba, Y.; Sasatani, M.; Nemoto, H.; Kato, A.; Adachi, I. *Tetrahedron* **2004**, *60*, 8199–8205. (b) Takahata, H.; Banba, Y.; Ouchi, H.; Nemoto, H. *Org. Lett.* **2003**, *5*, 2527–2529. (c) Takahata, H.; Banba, Y.; Ouchi, H.; Nemoto, H.; Kato, A.; Adachi, I. *J. Org. Chem.* **2003**, *68*, 3603–3607. (d) Banba, Y.; Abe, C.; Nemoto, H.; Kato, A.; Adachi, I.; Takahata, H. *Tetrahedron: Asymmetry* **2001**, *12*, 817–819.



FIGURE 2. Structures of a common building block 3 and 1-azasugars 4.

SCHEME 1^a



 a Reagents and conditions: (a) (i) cat. H₂O, 100 °C, (ii) Boc₂O, NaOH; (b) Grubbs' catalyst, CH₂Cl₂, rt.

We report herein a full paper¹³ describing the synthesis of both enantiomers of a variety of the 1-azasugars **4** such as isofagomine (**2**) by a general route starting from the (chiral) *N*-Boc-5-hydroxy-3-piperidene (**3**) as depicted in Figure 2.

Results and Discussion

Synthesis of racemic *N*-Boc-5-hydroxy-3-piperidene (**3**) as the starting material began with the regioselective opening of butadiene monoxide with allylamine followed by protection of the secondary amine to afford the metathesis precursor **5** (66%) together with the undesired ring-opening product **6** (20%).¹⁴ The Grubbs' catalyst¹⁵ could be used directly on **5** to afford the ring-closing metathesis product **3** in 99% yield (Scheme 1).

Ogasawara reported¹⁶ on the lipase-catalyzed transesterification of *N*-Cbz-5-hydroxy-3-piperidene with vinyl acetate. We applied this method to **3** using lipase and vinyl acetate. Of the various lipases tested, resolution was best achieved with lipase PS (*Pseudomonas cepacia*), immobilized on ceramic particles, in *tert*-butyl methyl ether at 40 °C, which gave the acetate (-)-**7** in 49% yield, along with the unreacted alcohol (+)-**3**, in 48% yield. In addition, the enzymatic hydrolysis of the acetate (-)-**7** with the same lipase in 0.1 M phosphate buffer afforded the enantiometic alcohol (-)-**3** in 98% yield. The enantiomeric purities of (+)- and (-)-**3** were >99% ee, as determined by chiral HPLC analysis after replacing the *N*-protecting group with Cbz (Scheme 2).

Alcohol (+)-3 was initially converted into the TBDPS derivative 8, thus avoiding the possible assistance of the hydroxyl group in the favored approach of the oxidant.

(15) Grubbs, R. H.; Chang, S. *Tetrahedron* 1998, 54, 4413–4450.
(16) Sakagami, H.; Ogasawara, K. Synthesis 2000, 521–524.

^{(10) (}a) Jespersen, T. M.; Dong, W.; Sierks, M. R.; Skrydstrup, T.; Lundt, I.; Bols, M. Angew. Chem., Int. Ed. Engl. 1994, 33, 1778-1779.
(b) Ichikawa, M.; Igarashi, Y.; Ichikawa, Y. Tetrahedron Lett. 1995, 36, 1767-1770. (c) Ichikawa, Y.; Igarashi, Y. Tetrahedron Lett. 1995, 36, 4585-4586. (d) Hansen, A.; Tagmose, T. M.; Bols, M. Chem. Commun. 1996, 2649-2650. (e) Nishimura, Y.; Satoh, T.; Kudo, T.; Kondo, S.; Takeuchi, T. Bioorg. Med. Chem. 1996, 4, 91-96. (f) Igarashi, Y.; Ichikawa, M.; Ichikawa, Y. Tetrahedron Lett. 1996, 37, 2707-2708. (g) Igarashi, Y.; Ichikawa, M.; Ichikawa, Y. Bioorg. Med. Chem. Lett. 1996, 6, 553-558. (h) Bols, M.; Hazell, R. G.; Thomsen, I. B. Chem. Eur. J. 1997, 3, 940-947. (i) Ichikawa, Y.; Igarashi, Y.; Ichikawa, M.; Suhara, Y. J. Am. Chem. Soc. 1998, 120, 3007-3018. (j) Nishimura, Y.; Shitara, E.; Adachi, H.; Toyoshima, M.; Nakajima, M.; Okami, Y.; Takeuchi, T. J. Org. Chem. 2000, 65, 2-11. (k) Shitara, E.; Nishimura, Y.; Kojima, F.; Takeuchi, T. Bioorg. Med. Chem. 2000, 8, 343-352. (l) Søhoel, H.; Liang, X.; Bols, M. J. Chem. Soc., Perkin Trans. 1 2001, 1584-1585. (m) Jakobsen, P.; Lundbeck, J. M.; Kristiansen, M.; Breinholt, J.; Demuth, H.; Pawlas, J.; Candela, M. P. T.; Andersen, B.; Westergaard, N.; Lundgren, K.; Asano, N. Bioorg. Med. Chem. 2001, 9, 733-744. (n) Liu, H.; Liang, X.; Søhoel, H. H.; Bülow, A.; Bols, M. J. Am. Chem. Soc. 2001, 123, 5116-5117. (11) (a) Kazmaier, U.; Schneider, C. Tetrahedron Lett. 1998, 39, 817-818. (b) Kim, Y. J.; Ichikawa M.; Ichikawa, Y. J. Org. Chem. 2000, 65, 2599-2602. (c) Mehta G.; Mohal N. Tetrahedron Lett. 1998, 39,

⁽¹³⁾ For a preliminary communication of this part of this work, see: Ouchi, H.; Mihara, Y.; Watanabe, H.; Takahata, H. *Tetrahedron Lett.* **2004**, *45*, 7053–7056.

^{(14) (}a) Evans, D. A.; Campos, K. R.; Tedrow, J. S.; Michael, F. E.; Gagné, M. R. J. Am. Chem. Soc. **2000**, 122, 7905–7920. (b) Baker, S. R.; Cases, M.; Keenan, M.; Lewis, R. A.; Tan, P. *Tetrahedron Lett.* **2003**, 44, 2995–2999. These papers reported the highly regioselective opening took place to provide only **5** as a single product in 52% and 39% yields, respectively.



^a Reagents and conditions: (a) vinyl acetate, lipase PS (immobilized on ceramic particles)/t-BuOMe, 40 °C, 18 h; (b) lipase PS (immobilized on ceramic particles)/0.1 M phosphate buffer (pH 7), acetone, 40 °C, 18 h.

SCHEME 3^a



^a Reagents and conditions: (a) TBDPSCl, imidazole, cat. DMAP/ CH₂Cl₂, rt, 3 h; (b) Oxone, aq Na₂EDTA, NaHCO₃, CF₃COCH₃/ CH₃CN, 0 °C, overnight.

Treatment of (+)-**3** with *tert*-butyldiphenylsilyl chloride under basic conditions gave the TBDPS derivative **8** in 99% yield. In an initial experiment, the oxidation of **8** with *m*-CPBA resulted in a low diastereoselectivity to give the anti epoxide **9** and the syn epoxide **10** in 41% and 23% yields, respectively. Subsequently, the use of (trifluoromethyl)methyldioxirane instead of *m*-CPBA as an oxidizing agent improved both the yield and the diastereoselectivity. Thus, the dioxirane, generated in situ¹⁷ from Oxone with 1,1,1-trifluoroacatone, was reacted with **8** to give the anti epoxide **9** and the syn epoxide **10** in 72% and 17% yields, respectively.

In diastereoselective epoxidation of allylically substituted six-membered cyclic alkene, dioxrane can be an oxygen atom to the olefin double bond of substrate from both faces, so there are two possible spiro transition states based on steric considerations.¹⁸ In the favored TS leading to anti epoxide **9**, there are little interactions between the R_1 and the allylic hydrogen of substrate. On the other hand, severe steric interaction between the R_1 and the allylic large OTBDPS group is expected in the disfavored TS leading to syn epoxide **10**. Therefore, it appears that an attack on the dioxirane preferably occurred preferentially at the less hindered anti side to the large OTBDPS group (Scheme 4).

The nucleophilic opening of the anti epoxide **9** with "higher order" cuprate¹⁹ free halide ions in the presence of boron trifluoride etherate as the activating species was carried out as follows: treatment of **9** with $(CH_2=CH)_2$ -

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SCHEME 4. Stereochemistry of the Epoxidation of 8







CuCNLi₂ in the presence of $BF_3 \cdot OEt_2$ at $-78 \degree C$ for 2 h gave 11 as the sole isolable product in 74% yield. Analogously, the reaction of 9 with Me₂CuCNLi₂ afforded only 12 in 71% yield. An attempt to employ Grignard reagents in the presence of cuprous bromide²⁰ resulted in no reaction. Although the rationale for this high selectivity remains unclear, the following mechanism is consistent with the results. The regiochemistry of the nucleophilic opening of the epoxide on a six-membered ring is mainly subject to trans diaxial opening (Fürst-Plattner rule).²¹ Consequently, a high regioselectivity would result if the opening proceeded through only one of the two possible half-chair conformations (A and B). Thus, the exclusive attack of the nucleophile at C-5 through conformer A would occur with trans diaxial opening. On the other hand, an attack of a nucleophile at C-4 of 9 through conformer B would be subject to steric hindrance by the pseudoequatorial OTBDPS group at C-3. Therefore, a trans diaxial opening through one of the half-chair conformers, namely conformer A, would occur (Scheme 5). A similar regioselectivity has been reported for ring-opening reactions of trans-3-(benzyloxy)-1,2-epoxycyclohexane derivatives.²²

With the vinyl product 11 in hand, our interest was directed to its conversion to isofagomine (2) and homo-

⁽¹⁷⁾ Yang, D.; Wong, M.-K.; Yip, Y.-C. J. Org. Chem. **1995**, 60, 3887–3889.

^{(18) (}a) Murray, R. W.; Singh, M.; Williams, B. L.; Moncrieff, H. M. J. Org. Chem. **1996**, 61, 1830–1841. (b) Yang, D.; Jiao, G.-S.; Yip, Y.-C.; Wong, M.-K. J. Org. Chem. **1999**, 64, 1635–1639.

⁽¹⁹⁾ Review: Lipshutz, B. H.; Wilhelm, R. S.; Kozlowski, J. A. Tetrahedron **1984**, 40, 5005–5038.

^{(20) (}a) Tius, M. A.; Fauq, A. H. J. Org. Chem. **1983**, 48, 4131–4132. (b) Roush, W. R.; Adam, M. A.; Walts, A. E.; Harris, D. J. J. Am. Chem. Soc. **1986**, 108, 3422–3434.

⁽²¹⁾ Eliel, E. L.; Wilen, S. H. Stereochemistry of Organic Compounds; Wiley-Interscience: New York, 1994; p 730.

^{(22) (}a) Crotti, P.; Renzi, G.; Favero, L.; Roselli, G.; Bussolo, V. D.;
Caselli, M. Tetrahedron 2003, 59, 1453-1467. (b) Crotti, P.; Bussolo,
V. D.; Favero, L.; Macchia, F.; Renzi, G.; Roselli, G. Tetrahedron 2002, 58, 7119-7133. (c) Crotti, P.; Bussolo, V. D.; Favero, L.; Pineschi, M.;
Marianucci, F.; Renzi, G.; Amici, G.; Roselli, G. Tetrahedron 2000, 56, 7513-7524. (d) Calvani, F.; Crotti, P.; Gardelli, C.; Pineschi, M. Tetrahedron 1994, 50, 12999-13022. (e) Chini, M.; Crotti, P.; Gardell, C.; Macchia, F. J. Org. Chem. 1994, 59, 4131-4137. (f) Montchamp, J.-L.; Migaud, M. E.; Frost, W. J. Org. Chem. 1993, 58, 7679-7684.

SCHEME 6^a



^a Reagents and conditions: (a) $(CH_2=CH)_2CuCNLi_2$ (5 equiv), BF₃·OEt₂ (2 equiv)/Et₂O, -78 °C, 2 h; (b) Me₂CuCNLi₂ (5 equiv), BF₃·OEt₂ (2 equiv)/Et₂O, -78 °C; (c) (i) cat. OsO₄, NaIO₄/50% EtOH, rt, overnight, (ii) NaBH₄/50% EtOH, rt, 1 h, (iii) 10% HCl/ dioxane, reflux, 1 h; (d) (i) 9-BBN/THF, rt, 6 h, (ii) H₂O₂, 3 mol/L of NaOH/THF, rt, 1 h, (iii) 10% HCl/dioxane, reflux, 1 h; (e) 10% HCl/dioxane, reflux, 1 h.



FIGURE 3. Structures of the enantiomers of 2, 13, and 14,



FIGURE 4. Structures of three 3,4,5-trihydroxypiperidines isolated from *Eupatorium fortunei* TURZ.

isofagomine (13). Oxidative cleavage of the vinyl group of 11 with OsO₄ and NaIO₄ afforded the aldehyde, which without purification, and after reduction with NaBH₄ followed by deprotection, afforded isofagomine (2) in 85% combined yield. The optical rotation and spectral characteristics of 2 were in good agreement with those reported in the literature.^{11g} Next, the hydroboration of the vinyl group of **11** with 9-BBN followed by treatment with hydrogen peroxide gave the corresponding primary alcohol. Deprotection of the alcohol with 10% HCl in 1,4dioxane afforded 13 in 86% combined yield. Conversion of 12 to 5'-deoxyisofagomine (14) was accomplished by deprotection. The complete deprotection of 12 by treatment with 10% HCl in 1,4-dioxane afforded 14 in 92% yield. Thus the first synthesis of 13 and 14 was achieved (Scheme 6).

Enantiomers of 2, 13, 14, and 15-17 were also prepared starting from (-)-3, following the same procedure as above (Figure 3).

The above successes led us to consider the synthesis of some of the most important 3,4,5-piperidine triols, namely, **18**, **19**, and **20** (Figure 4). These trihydroxy piperidines are regarded as derivatives of their parent deoxynojirimycin and have been shown to possess moderate to good glycosidase inhibitory activity. A careful





 a Reagents and conditions: (a) (i) cat. OsO4, NMO, acetone, (ii) TBAF, THF, rt; (b) (i) 10% HCl, dioxane, reflux, (ii) Amberlite IRA-410.

consideration of these structural moieties reveals that these molecules can also be classified as 1-*N*-iminosugartype glycosidase inhibitors, although they do not resemble any of the presently known pyranose sugars.

Triols 18, 19, and 20 were isolated from *Eupatorium* fortunei TURZ by Kusano and co-workers²³ in 1995 and have been shown to be the active components of the extracts of this plant, which are traditionally used in Chinese and Japanese folk medicine as a diuretic, antipyretic, emmenagogue, and antidiabetic agent. Triols 18-20 have also been synthesized by Ganem²⁴ and others²⁵ and have been shown to be good selective inhibitors of glycosidases.

We realized that our precursors (+)-, (-)-, and (\pm) -3 would be ideal candidates for the synthesis of these molecules together with the enantiomer of 18, 21. The synthesis of des(hydroxymethyl)deoxymannojirimycin (18) commenced with ent-8, which was previously synthesized from (-)-3. The stereoselective dihydroxylation of the double bond was also examined. Treatment of ent-8 with a catalytic amount of OsO₄ (5 mol %) and 4-methylmorphorine N-oxide as a cooxidant²⁶ gave an inseparable mixture of diastereomeric diols, which was subjected to treatment with TBAF to provide the triol **22** as a major product along with minor amounts of triol 23. This high diastereoselectivity in the dihydroxylation step can be attributed to the a steric blocking of an allylic O-TBDPS substituent.^{25h} Deprotection of $\mathbf{22}$ with HCl in dioxane followed by treatment with an ion-exchange resin (Amberlite IRA-410 OH⁻ form) afforded 18 in 98% combined yield. In a similar procedure, the enantiomer **21** was prepared from (+)-**3** in 69% yield (Scheme 7). The spectral data and optical rotation for 18 and 21 were found to be in good agreement with reported values from the literature.^{23,25c}Donohoe reported²⁷ that osmium tetroxide produces a bidendate and reactive complex with TMEDA, which can be used to achieve the directed

⁽²³⁾ Sekioka, T.; Shibano, M.; Kusano, G. Nat. Med. (Tokyo, Jpn.) **1995**, 49, 332–335.

⁽²⁴⁾ Bernotas, R. C.; Papandreou, G.; Urbach, J.; Ganem, B. Tetrahedron Lett. **1990**, *31*, 3393–3396. (25) (a) Tschamber, T.; Backenstrass, F.; Neuburger, M.; Zehnder,

^{(25) (}a) Tschamber, T.; Backenstrass, F.; Neuburger, M.; Zehnder, M.; Streith, J. Tetrahedron 1994, 50, 1135–1152. (b) Legler, G.; Stütz, A. E.; Immich, H. Carbohydr. Res. 1995, 272, 17–30. (c) Godskesen, M.; Lundt, I.; Madsen, R.; Winchester, B. Bioorg. Med. Chem. 1996, 4, 1857–1865. (d) Sames, D.; Polt, R. Synlett 1995, 552–554. (e) Amat, M.; Llor, N.; Huguet, M.; Molins, E.; Espinosa, E.; Bosch, J. Org. Lett. 2001, 3, 3257–3260. (f) Patil, N. T.; John, S.; Sabharwal, S. G.; Dhawale, D. D. Bioorg. Med. Chem. 2002, 10, 2155–2160. (g) Pandey, G.; Kapur, M. Org. Lett. 2002, 4, 3883–3886. (h) Han, H. Tetrahedron Lett. 2003, 44, 1567–1569. In this paper, a highly stereocontrolled synthesis of 18 and 20 as hydrochloride salts from the O-MOM protected piperidene of 3 has been reported. (i) Pandey, G.; Kapur, M. S.; Gaikwad, S. M. Org. Biomol. Chem. 2003, 1, 3321–3326.

⁽²⁶⁾ VanRheenen, V.; Kelly, R. C.; Cha, D. Y. Tetrahedron Lett. 1976, 17, 1973–1976.

SCHEME 8^a



 a Reagents and conditions: (a) (i) OsO4,TMEDA, CH₂Cl₂, -78 °C, (ii) conc HCl, MeOH, rt, (iii) Boc₂O, Et₃N, MeOH; (b) (i) 10% HCl, dioxane, reflux, (ii) Amberlite IRA-410; (c) (i) 10% H₂SO₄, dioxane, 95 °C, 3 h, (ii) Amberlite IRA-410.

dihydroxylation of cyclic allylic alcohols. Under this condition, hydrogen bonding control leads to the formation of the syn isomer in almost every case. Therefore, the oxidation of (\pm) -3 with a combination of OsO₄ with TMEDA gave the *all-cis*-triol **23** and (\pm) -**22** in moderate selectivity (4:1) in 66% and 16% yields, respectively. Finally, removal of the Boc group from 23, followed by ion-exchange chromatographic purification yielded 19 in 94% yield, the spectral data for which were in excellent agreement with data reported in the literature. Next, the anti epoxide (\pm) -9 obtained from (\pm) -8 according to the same procedure for the preparation of (+)-9 was hydrolyzed with 10% H₂SO₄ in dioxane to afford the triol 20 as a major product and (\pm) -18 as a minor one in 55% and 20% yields, respectively. This moderate diastereoselectivity compared with the high selectivity of nucleophilic cleavage of 9 with vinylmetal may be due to the following reason. Because partial desilylation prior to ring opening of epoxide in acidic condition can occur, the steric hindrance between nucleophile (H₂O) and allvlic hydroxy substituent in analogous conformer B described in Scheme 5 would diminish.

Summary

An efficient synthesis of isofagomine (2), homoisofagomine (13), 5-deoxyisofagomine (14), and their enantiomers 15-17 with a nitrogen atom at the anomeric position was achieved in 44-50% overall yields from the readily obtainable chiral *N*-Boc-5-hydroxy-3-piperidene (3). The reaction involved the epoxidation into a double bond and a regioselective epoxide ring opening reaction with higher ordered cuprates. To our knowledge our route to isofagomine (2) is shorter and more efficient than the other reported syntheses.¹⁰ In addition, the stereoselective synthesis of all four of the stereoisomers of 3,4,5-trihydroxypiperidines (18-21) was achieved from the common building block 3. This synthetic route permits the preparation of substantial amounts of 1-azasugars having a glucose configuration and would be suitable for

further studies of such compounds as glycosidase inhibitors.

Experimental Section

tert-Butyl N-Allyl-N-(2-hydroxy-3-butenyl)carbamate (5) and tert-Butyl Allyl-1-hydroxybut-3-enylcarbamate (6). To a solution of allylamine (8.5 mL, 20 mmol) and water $(530 \ \mu\text{L})$ was added butadiene monoxide $(3.0 \ \text{mL}, 37.5 \ \text{mmol})$ at 15 °C. The mixture was warmed to 100 °C and stirred for 6 h. The reaction mixture was concentrated under reduced pressure. The residue was dissolved in dioxane (50 mL) and water (10 mL), 1 mol/L of NaOH (40 mL, 40 mmol), and (Boc)₂O (8.37 g, 40 mmol) were added. The mixture was stirred at room temperature overnight. The residue obtained after evaporation was taken up in Et_2O (200 mL), and washed with water (100 mL), 20% citric acid (100 mL), and brine (100 mL). The combined aqueous solutions were extracted with $Et_2O(2)$ \times 50 mL). The combined organic solutions were dried over MgSO₄, filtered, concentrated in vacuo, and purified by flash column chromatography on silica gel (n-hexane:AcOEt = 4:1-1:1) to give 5 (5.62 g, 66%) as a colorless oil and 6 (1.68 g, 20%) as a colorless oil.

5: ¹H NMR (600 MHz, CDCl₃) δ 1.44 (s, 9H), 3.26 (br s, 1H), 3.31 (br s, 1H), 3.66 (br s, 1H), 3.74–3.94 (m, 2H), 4.29–4.32 (m, 1H), 5.09–5.15 (m, 3H), 5.32 (d, J = 16.5 Hz, 1H), 5.66–5.84 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 28.3, 51.7, 53.4, 72.7, 80.4, 115.7, 116.4, 133.8, 138.4, 157.6. IR (neat) (cm⁻¹) 3432, 1675. EI-MS (*m/z*) 228 (M⁺ + 1). HRMS calcd for C₁₀H₁₇NO₃ 227.1521, found 227.1538. Anal. Calcd for C₁₂H₂₁-NO₃: C, 63.41; H, 9.31; N, 6.16. Found: C, 63.10; H, 9.52; N, 6.28.

6: ¹H NMR (400 MHz, CDCl₃) δ 1.46 (s, 9H), 2.27–3.00 (br, 1H), 3.77 (br d, J = 7.1 Hz, 4H), 4.38 (br d, J = 5.6 Hz, 1H), 5.06–5.26 (m, 4H), 5.74–5.96 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 28.4, 48.3, 61.1, 63.6, 80.3, 117.5, 134.0, 135.3, 156.3. IR (neat) (cm⁻¹) 3436, 1694. EI-MS (m/z) 228 (M⁺ + 1). HRMS calcd for C₁₂H₂₁NO₃ 227.1521, found 227.1546.

 (\pm) -N-tert-Butoxycarbonyl-5-hydroxy-3-piperidene ((\pm)-**3).** To a deoxygenated solution of **5** (4.54 g, 20 mmol) in CH₂-Cl₂ (340 mL) under argon atomosphere was added bis-(tricyclohexylphosphine)benzylidine ruthenium(IV) dichloride (0.58 g, 0.71 mmol, 5 mol %). The solution was stirred at room temperature overnight, then concentrated under reduced pressure, and the residue was purified by flash column chromatography on silica gel (*n*-hexane:AcOEt = 2:1) to give (±)-3 (3.95 g, 99%) as colorless prisms; mp 55–56 °C (nhexane). ¹H NMR (600 MHz, CDCl₃) δ 1.47 (s, 9H), 1.64 (br s, 1H), 3.53 (br s, 1H), 3.58 (br s, 1H), 3.80 (dd, J = 18.7, 2.2 Hz, 1H), 3.97 (br s, 1H), 4.19 (br s, 1H), 5.84 (br s, 1H), 5.89-5.93 (m, 1H).¹³C NMR (67.8 MHz, CDCl₃) δ 28.4, 43.3, 47.7, 63.6, 80.1, 126.9, 128.3, 155.0. IR (KBr) (cm⁻¹) 3255, 1699. EI-MS (m/z) 199 (M⁺). HRMS calcd for C₁₀H₁₇NO₃ 199.1169, found 199.1208. Anal. Calcd for C₁₀H₁₇NO₃: C, 60.28; H, 8.60; N, 7.03. Found: C, 60.46; H, 8.50; N, 6.87.

(S)-N-tert-Butoxycarbonyl-5-hydroxy-3-piperidene ((+)-3) and (R)-N-tert-Butoxycarbonyl-5-acetoxy-3-piperidene ((-)-7). A mixture of (\pm) -3 (1.99 g, 10 mmol) and vinyl acetate (4.3 g, 50 mmol) in tert-butyl methyl ether (100 mL) was stirred with lipase (1 g, immobilized on Ceramic particles from *Psedomonas cepacia*) at 40 °C for 18 h. After filtration, the filtrate was evaporated under reduced pressure. The residue was purified by flash column chromatography on silica gel to give acetate (-)-7 (1.18 g, 49%) as a colorless oil from the *n*-hexane/AcOEt (3:1) fraction and the alcohol (+)-3 (0.96 g, 48%) as colorless needles from the *n*-hexane/AcOEt (1:1) fraction.

(+)-3: mp 70–71 °C (*n*-hexane). $[\alpha]^{32}_{D}$ +83.7 (*c* 1.08, CHCl₃). (–)-7: $[\alpha]^{30}_{D}$ –110.4 (*c* 2.20, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ 1.46 (s, 9H), 2.05 (s, 3H), 3.47 (dd, J = 13.9, 3.7 Hz, 1 H), 3.76 (d, J = 19.4, 1H), 3.76 (dd, J = 13.9, 4.0 Hz, 1H), 4.12 (m, 1H), 5.15 (br s, 1H), 5.85–5.97 (m, 2H). ¹³C NMR (67.8)

^{(27) (}a) Donohoe, T. J.; Blades, K.; Moore, P. R.; Waring, M. J.; Winter, J. J. G.; Helliwell, M.; Newcombe, Ni. J.; Stemp, G. J. Org. Chem. 2002, 67, 7946-7956. (b) Donohoe, T. J. Synlett 2002, 1223-1232.

MHz, CDCl₃) δ 21.1, 28.4, 42.8, 44.9, 65.7, 79.9, 123.7, 130.4, 154.5, 170.3. IR (neat) (cm⁻¹) 1740, 1700. EI-MS (*m/z*) 242 (M⁺ + 1). HRMS calcd for C₁₂H₁₉NO₄ 241.1314, found 241.1321.

(*R*)-*N*-tert-Butoxycarbonyl-5-hydroxy-3-piperidene ((-)-3). A mixture of acetate (-)-7 (1.1 g, 4.5 mmol) and acetone (3.14 mL) in phosphate buffer (0.1 mol/L, pH 7.0, 51 mL) was stirred with lipase (0.46 g, immobilized on Ceramic particles from *Psedomonas cepacia*) at 40 °C for 18 h. After filtration, the filtrate was extracted with AcOEt (3×50 mL). The combined organic extracts were washed with brine (50 mL), dried over MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (*n*-hexane:AcOEt = 1:1) to give (-)-3 (0.88 g, 98%) as colorless needles; mp 71-72 °C (*n*-hexane). [α]³⁰_D -83.3 (*c* 1.10, CHCl₃).

(S)-N-tert-Butoxycarbonyl-5-(tert-butyldiphenylsilyloxy)-3-piperidene (8). To a solution of (+)-3 (996 mg, 5 mmol) in CH₂Cl₂ (10 mL) was added imidazole (511 mg, 7.5 mmol), DMAP (122 mg, 1 mmol), and tert-butylchlorodiphenylsilane (1.512 g, 5.5 mmol). The mixture was stirred at room temperature for 3 h. The reaction mixture was filtered through a Celite pad. The filtrate was washed with brine (10 mL), dried over MgSO₄, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (*n*-hexane:AcOEt = 8:1) to give 8 (2.35 g, 99%) as a colorless oil; $[\alpha]^{27}_{D}$ +22.6 (c 1.80, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ 1.08 (s, 9H), 1.40 (s, 9H), 3.21 (br s, 1H), 3.62-3.96 (m, 3H), 4.25 (br s, 1H), 5.58-5.72 (m, 2H), 7.36-7.44 (m, 6H), 7.67-7.70 (m, 4H). $^{13}\mathrm{C}$ NMR (67.8 MHz, CDCl₃) δ 19.2, 26.9, 28.4, 42.7, 47.9, 65.1, 79.5, 126.0, 127.4, 129.2, 129.5, 133.7, 135.5, 154.4. IR (neat) (cm⁻¹) 1703. EI-MS (m/z) 438 (M⁺ + 1). HRMS calcd for C₂₆H₃₅NO₃Si 437.2386, found 437.2448. Anal. Calcd for C₂₆H₃₅NO₃Si: C, 71.35; H, 8.06; N, 3.20. Found: C, 71.16; H, 7.96; N, 3.23.

(3R,4R,5R)-N-tert-Butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4,5-epoxypiperidine (9) and (3R,4S,5S)-*N-tert*-Butoxycarbonyl-3-(*tert*-butyldiphenylsilyloxy)-4,5-epoxypiperidine (10). To a cooled (0 °C) solution of 8 (945 mg, 2.16 mmol) in CH₃CN (20 mL) was added 4 mmol/L of aq Na₂EDTA (10 mL, 0.04 mmol) and 1,1,1-trifluoroacetone (2 mL). The mixture of NaHCO₃ (1.26 g, 15 mmol) and Oxone (6.13 g, 10 mmol) as a solid was added slowly over a period of 1 h at 0 °C. After being stirred at 0 °C overnight, the solution was quenched by adding water (60 mL) and extracted with $CHCl_3$ (3 × 60 mL). The organic extracts were combined, dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (*n*-hexane:AcOEt = 8:1) to give 9 (706 mg, 72%) as colorless prisms and 10 (170 mg, 17%) as colorless needles.

9: mp 70–72 °C (*n*-pentane); $[\alpha]^{30}{}_{\rm D}$ +29.2 (*c* 1.11, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.10 (s, 9H), 1.37 and 1.46 (2 s, 9H), 3.07–3.41 (m, 4H), 3.07–3.88 (m, 2H), 4.09–4.15 (m, 1H), 7.36–7.45 (m, 6H), 7.63–7.69 (m, 4H). ¹³C NMR (67.8 MHz, CDCl₃) δ 19.3, 27.0, 28.4, 41.4, 45.3, 50.6, 53.7, 65.2, 79.8, 127.7, 129.8, 133.1, 135.5, 154.9. IR (KBr) (cm⁻¹) 1701. EI-MS (*m/z*) 453 (M⁺). HRMS calcd for C₂₆H₃₅NO₄Si 453.2336, found 453.2343. Anal. Calcd for C₂₆H₃₅NO₄Si: C, 68.84; H, 7.78; N, 3.09. Found: C, 69.03; H, 7.64; N, 2.93.

10: mp 110–111 °C (*n*-pentane); $[\alpha]^{26}{}_{\rm D}$ –6.2 (*c* 0.80, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.09 (s, 9H), 1.35 (s, 9H), 2.87 (br t, J = 10.7 Hz, 1H), 3.09 (br s, 1H), 3.20–3.40 (m, 2H), 3.75 (br s, 1H), 3.99–4.03 (m, 2H), 7.37–7.46 (m, 6H), 7.67– 7.74 (m, 4H). ¹³C NMR (67.8 MHz, CDCl₃) δ 19.3, 27.0, 28.3, 40.6, 43.1, 53.3, 54.6, 67.5, 79.9, 127.7, 129.8, 133.1, 135.6, 154.0. IR (KBr) (cm⁻¹) 1698. EI-MS (*m/z*) 454 (M⁺ + 1). HRMS calcd for C₂₆H₃₅NO₄Si 453.2336, found 453.2351. Anal. Calcd for C₂₆H₃₅NO₄Si: C, 68.84; H, 7.78; N, 3.09. Found: C, 68.96; H, 7.88; N, 3.12.

(3R,4R,5S)-N-tert-Butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4-hydroxy-5-vinylpiperidine (11). A solution of *n*-BuLi (3.125 mL, 1.6 M in *n*-hexane, 5 mmol) was added to a solution of tetravinyltin (1.14 g, 5 mmol) in Et₂O (7 mL) at room temperature and the mixture was stirred for 1 h at the same temperature. The solution was added to a suspension of CuCN (224 mg, 2.5 mmol) in dry Et₂O (20 mL) at -78 °C and the mixture was stirred under argon atmosphere at -78 °C for 10 min. The resulting solution was warmed to -10 °C and stirred for 30 min. The reaction mixture was cooled to -78 °C and a solution of 9 (454 mg, 1 mmol) in dry Et₂O (20 mL) was added to the reaction mixture dropwise via syringe at -78 °C. The mixture was stirred at -78 °C for 10 min. BF₃·OEt₂ (4 mol/L in Et₂O, 5 mL, 2 mmol) was added to the reaction mixture at -78 °C and the solution was stirred for 2 h. The reaction mixture was quenched with 28% NH₄-OH/sat. NH₄Cl (1:9) solution (50 mL) and stirred overnight at room temperature. The organic layer was separated and the aqueous layer was extracted with Et_2O (2 \times 20 mL). The combined organic phases were washed with brine (20 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (*n*-hexane:AcOEt = 8:1) to give 11 (357) mg, 74%) as a colorless oil; $[\alpha]^{24}$ _D -10.5 (c 1.16, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ 1.09 (s, 9H), 1.31 (s, 9H), 2.12 (br s, 1H), 2.23 (br s, 1H), 2.50 (br s, 1H), 2.65 (dd, J = 11.0, 12.5Hz, 1H), 3.42 (t, J = 9.0 Hz, 1H), 3.51–3.55 (m, 1H), 3.82– 4.23 (m, 2H), 5.16-5.20 (m, 2H), 5.66-5.72 (m, 1H), 7.38-7.45 (m, 6H), 7.68–7.71 (m, 4H). ¹³C NMR (67.8 MHz, CDCl₃) $\delta \ 19.4, 27.0, 28.3, 45.8, 46.5, 48.7, 73.6, 77.7, 79.8, 117.7, 127.7,$ 129.8, 133.3, 135.7, 135.9, 154.0. IR (neat) (cm⁻¹) 3472, 1698. EI-MS (m/z) 482 $(M^+ + 1)$. HRMS calcd for C₂₈H₃₉NO₄Si 481.2649, found 481.2686. Anal. Calcd for $\mathrm{C}_{28}\mathrm{H}_{39}\mathrm{NO}_4\mathrm{Si:}$ C, 69.82; H, 8.16; N, 2.91. Found: C, 70.10; H, 8.07; N, 2.77.

(3R,4R,5R)-3,4-Dihydroxy-5-(hydroxymethyl)piperidine (2, isofagomine). To a solution of 11 (193 mg, 0.4 mmol) in EtOH (4 mL) and H₂O (4 mL) was added an aqueous solution of 4% OsO_4 (131 μ L, 0.02 mmol) and the solution was stirred at room temperature for 10 min. NaIO₄ (190 mg, 0.88 mmol) as a powder was added slowly over a period of 30 min, and the mixture was stirred at room temperature for 24 h. NaBH₄ (76 mg, 2 mmol) as a powder was added, and the reaction mixture was stirred at room temperature for 1 h. The solvent was removed in vacuo. The residue was diluted with CHCl₃ (80 mL) and successively washed with 10% Na₂S₂O₃ (20 mL), sat. NaHCO₃ (20 mL), and brine (10 mL). The organic solution was dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (n-hexane:AcOEt = 1:1)to give (3R,4R,5R)-N-tert-butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4-hydroxy-5-(hydroxymethyl)piperidine (167 mg, 86%) as a colorless oil; $[\alpha]^{25}$ _D -15.9 (*c* 1.35, CHCl₃). ¹H NMR $(600 \text{ MHz}, \text{CDCl}_3) \delta 1.09 \text{ (s, 9H)}, 1.31 \text{ (s, 9H)}, 1.66 \text{ (br s, 1H)},$ 2.48 (br s, 3H), 2.63 (t, J = 11.0 Hz, 1H), 3.48–3.56 (m, 2H), 3.67 (s, 2H), 3.85-4.16 (m, 2H), 7.39-7.46 (m, 6H), 7.67-7.69 (m, 4H). ¹³C NMR (67.8 MHz, CDCl₃) δ 19.4, 27.0, 28.3, 42.8, 43.9, 48.5, 63.2, 73.6, 77.7, 79.9, 127.8, 129.9, 133.3, 135.5, 154.2. IR (neat) (cm⁻¹) 3420, 1696, 1674. EI-MS (m/z) 486 (M + H⁺). HRMS calcd for C₂₇H₃₉NO₅Si 485.2598, found 485.2585. To a solution of the prepared diol (160 mg, 0.33 mmol) in dioxane (2.5 mL) was added a solution of 10% HCl (10 mL), and the solution was refluxed for 1 h. After the reaction mixture was cooled to room temperature and diluted with H₂O (50 mL), the resulting dilute solution was washed with Et_2O $(2 \times 20 \text{ mL})$. The aqueous layer was concentrated under reduced pressure. The residue was dissolved in 28% NH₄OH and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (28% NH₄OH:EtOH = 1:10) to give 2 (48.5 mg, 99%) as a white powder; $[\alpha]^{25}{}_D$ +25.4 (c 1.30, EtOH). ¹H NMR (600 MHz, D_2O) δ 1.61–1.68 (m, 1H), 2.33-2.38 (m, 2H), 3.05 (dd, J = 13.0, 3.5 Hz, 1H), 3.10 (dd, J= 12.1, 4.8 Hz, 1H), 3.27 (t, J = 9.9 Hz, 1H), 3.43–3.48 (m, 1H), 3.59 (dd, J = 11.4, 7.0 Hz, 1H), 3.78 (dd, J = 11.4, 3.3 Hz, 1H).¹³C NMR (67.8 MHz, D_2O) δ 43.9, 45.7, 48.8, 59.8, 71.4, 73.1. EI-MS (m/z) 147 (M⁺). HRMS calcd for C₆H₁₃NO₃ 147.0895, found 147.0845.

(3R,4R,5S)-3,4-Dihydroxy-5-(hydroxyethyl)piperidine (13, homoisofagomine). To a cooled (0 °C) solution of 11 (96 mg, 0.2 mmol) in THF (3 mL) was added a solution of 9-BBN (0.5 mol/L in THF, 2 mL, 1 mmol). The mixture was warmed to room temperature and stirred for 6 h. The reaction mixture was then cooled to 0 °C. To this solution was added 3 mol/L of NaOH solution (1 mL) and 30% H₂O₂ solution (1 mL). The mixture was stirred at room temperature for 1 h. A sat. NH₄Cl solution (3 mL) was added, and the mixture was extracted with AcOEt (3×20 mL). The organic extracts were combined, dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (*n*-hexane:AcOEt = 1:1) to give (3R,4R,5S)-N-tert-butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4-hydroxy-5-(hydroxyethyl)piperidine (93 mg, 93%) as a colorless oil; [a]²⁵_D -4.97 (c 1.30, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ 1.08 (s, 9H), 1.29 (br s, 9H), 1.47–1.75 (m, 4H), 2.37 (br s, 1H), 2.61 (t, J = 11.7 Hz, 1H), 2.94 (br s, 1H), 3.33 (t, J)= 9.0 Hz, 1H), 3.47-3.51 (m, 1H), 3.59-3.63 (m, 1H), 3.69-3.73 (m, 1H), 3.84-4.20 (m, 2H), 7.38-7.51 (m, 6H), 7.66-7.69 (m, 4H). IR (neat) (cm⁻¹) 3323, 1697, 1673. EI-MS (m/z) 499 (M⁺). HRMS calcd for C₂₈H₄₁NO₅Si 499.2754, found 499.2740. To a solution of the above diol (180 mg, 0.36 mmol) in dioxane (2.5 mL) was added a solution of 10% HCl (10 mL), and the solution was heated at reflux for 1 h. After the reaction mixture was cooled to room temperature and diluted with H₂O (50 mL), the resulting dilute solution was washed with Et_2O $(2 \times 20 \text{ mL})$. The aqueous layer was concentrated under reduced pressure. The residue was dissolved in MeOH and made alkaline with Na₂CO₃. The mixture was filtered and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (10% NH₄OH:MeOH = 1:50) to give **13** (55 mg, 95%) as a white powder; $[\alpha]^{20}$ +17.70 (c 1.10, EtOH). ¹H NMR (600 MHz, D₂O) δ 1.22–1.28 (m, 1H), 1.36-1.43 (m, 1H), 1.76-1.82 (m, 1H), 2.11 (dd, J = 12.8, 11.7)Hz, 1H), 2.23 (dd, J = 12.3, 10.8 Hz, 1H), 2.87 (ddd, J = 13.0, 4.2, 1.5 Hz, 1H), 2.94-3.01 (m, 2H), 3.27-3.32 (m, 1H), 3.46-3.55 (m, 2H). $^{13}\!C$ NMR (67.8 MHz, $D_2O)$ δ 76.5, 71.3, 58.8, 48.8, 47.6, 39.1, 30.7. IR (neat) (cm⁻¹) 3324, 1697, 1674. EI-MS (m/ z) 161 (M⁺). HRMS calcd for $C_7H_{15}NO_3$ 161.1052, found 161.1088.

(3R,4R,5S)-N-tert-Butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4-hydroxy-5-methylpiperidine (12). To a suspension of CuCN (135 mg, 1.5 mmol) in dry Et₂O (15 mL) at -78 °C was added a solution of MeLi (2.9 mL, 1.04 mol/L in Et₂O, 3 mmol) at -78 °C and the mixture was stirred under argon atmosphere at -78 °C for 10 min. The resulting solution was warmed to -10 °C then stirred for 30 min. The reaction mixture was cooled to -78 °C, and a solution of 9 (273 mg, 0.6 mmol) in dry Et_2O (15 mL) was added to the reaction mixture dropwise via syringe at -78 °C. The mixture was stirred at -78 °C for 10 min. BF₃·OEt₂ (4 mol/L in Et₂O, 3 mL, 1.2 mmol) was added to the reaction mixture at -78 °C and the solution was stirred for 2 h. The reaction mixture was quenched with 28% NH₄OH/sat. NH₄Cl (1:9) solution (30 mL) and stirred overnight at room temperature. The organic layer was separated and the aqueous layer was extracted with Et₂O $(2 \times 30 \text{ mL})$. The combined organic phases were washed with brine (20 mL), dried over MgSO4, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (*n*-hexane:AcOEt = 8:1) to give 12 (200 mg, 71%) as a colorless oil; $[\alpha]^{23}_{D} - 16.7$ (c 1.25, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ 0.97 (d, J = 6.6 Hz, 3H), 1.09 (s, 9H), 1.30 (s, 9H), 1.43 (br, 1H), 2.15-2.36 (m, 2H), $2.62 \,(dd, J = 12.5, 11.0 \text{ Hz}, 1\text{H}), 3.18 \,(t, J = 9.0 \text{ Hz}, 1\text{H}), 3.46-$ 3.51 (m, 1H), 3.97 (br, 2H), 7.37-7.46 (m, 6H), 7.65-7.73 (m, 4H). ¹³C NMR (67.8 MHz, CDCl₃) δ 14.6, 19.3, 26.9, 28.2, 35.9, 48.7, 74.0, 79.6, 80.2, 127.9, 129.9, 133.3, 135.6, 154.2. IR (neat) (cm^{-1}) 3476, 1699. EI-MS (m/z) 470 $(M^+ + 1)$. HRMS calcd for C₂₇H₃₉NO₄Si 469.2649, found 469.2657.

(3R,4R,5S)-3,4-Dihydroxy-5-methylpiperidine (14, 5'deoxyisofagomine). To a solution of 12 (141 mg, 0.30 mmol) in dioxane (2.5 mL) was added a solution of 10% HCl (10 mL), and the solution was heated at reflux for 1 h. After the reaction mixture was cooled to room temperature and diluted with H₂O (50 mL), the resultant dilute solution was washed with Et_2O $(2 \times 20 \text{ mL})$. The aqueous layer was concentrated under reduced pressure. The residue was dissolved in MeOH and made alkaline with Na₂CO₃. The mixture was filtered and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (10% NH₄OH:MeOH = 1:50) to give 14 (37 mg, 92%) as a white powder; $[\alpha]^{25}_{D}$ +6.5 (c0.5, EtOH). ¹H NMR (600 MHz, D_2O) δ 0.81 (d, J = 6.6 Hz, 3H), 1.36–1.44 (m, 1H), 2.09 (dd, J = 12.8, 11.7 Hz, 1H), 2.25 (dd, J = 12.1, 11.0 Hz, 1H), 2.76 (ddd, J = 13.1, 4.4, 1.5 Hz)1H), 2.91 (t, J = 9.5 Hz, 1H), 2.97 (ddd, J = 12.3, 5.1, 1.6 Hz, 1H), 3.27–3.32 (m, 1H). $^{13}\mathrm{C}$ NMR (67.8 MHz, D2O) δ 78.1, 71.2, 49.7, 49.0, 36.8, 13.1. EI-MS (m/z) 131 (M⁺). HRMS calcd for C₆H₁₃NO₂ 131.0946, found 131.0984.

(*R*)-*N*-tert-Butoxycarbonyl-5-(tert-butyldiphenylsilyloxy)-3-piperidene (ent-8). This compound was prepared from (–)-3 (897 mg, 4.5 mmol), imidazole (511 mg, 7.5 mmol), DMAP (122 mg, 1 mmol), and *tert*-butylchlorodiphenylsilane (1.374 g, 5 mmol) according to the procedure described for the preparation of 8 to give ent-8 (1.96 g, 99%); $[\alpha]^{29}_{D}$ –22.3 (c 1.40, CHCl₃).

(3S,4S,5S)-*N*-tert-Butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4,5-epoxypiperidine (ent-9) and (3S,-4R,5R)-*N*-tert-Butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4,5-epoxypiperidine (ent-10). These compounds were prepared from ent-8 (1.40 g, 3.2 mmol), 4 mmol/L of aq Na₂-EDTA (17.5 mL, 0.07 mmol), 1,1,1-trifluoroacetone (3.5 mL), NaHCO₃ (2.21 g, 26.25 mmol), and Oxone (10.73 g, 17.5 mmol) according to the procedure described for the preparation of **9** and **10** to give ent-**9** (1.04 g, 72%) and ent-**10** (272 mg, 19%).

ent-9: $[\alpha]^{31}_{D}$ -29.7 (*c* 1.82, CHCl₃).

ent-10: $[\alpha]^{30}_{D}$ +6.9 (*c* 1.10, CHCl₃).

(3*S*,4*S*,5*R*)-*N*-*tert*-Butoxycarbonyl-3-(*tert*-butyldiphenylsilyloxy)-4-hydroxy-5-vinylpiperidine (*ent*-11). This compound was prepared from *ent*-9 (454 mg, 1 mmol), *n*-BuLi (3.125 mL, 1.6 M in *n*-hexane, 5 mmol), tetravinyltin (1.14 g, 5 mmol), CuCN (224 mg, 2.5 mmol), and BF₃·OEt₂ (4 mol/L in Et₂O, 5 mL, 2 mmol) according to the procedure described for the preparation of **11** to give *ent*-**11** (383 mg, 80%); $[\alpha]^{30}_{D}$ +10.6 (*c* 1.00, CHCl₃).

(3S,4S,5S)-3,4-Dihydroxy-5-(hydroxymethyl)piperidine (15). This compound was prepared from *ent*-11 (241 mg, 0.5 mmol), 4% OsO₄ (164 μ L, 0.025 mmol), NaIO₄ (238 mg, 1.1 mmol), and NaBH₄ (95 mg, 2.5 mmol) according to the procedure described for the preparation of 2 to give (3S,4S,5S)-*N*-tert-butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4-hydroxy-5-(hydroxymethyl)piperidine (216 mg, 89%); [α]³⁰D +15.5 (*c* 1.16, CHCl₃). This compound was prepared from *ent*-8 (131 mg, 0.27 mmol) according to the procedure described for the preparation of 2 to give 15 (38 mg, 96%); [α]²⁴D -23.8 (*c* 1.10, EtOH).

(3*S*,4*S*,5*R*)-3,4-Dihydroxy-5-(hydroxyethyl)piperidine (16). This compound was prepared from *ent*-11 (121 mg, 0.5 mmol), 9-BBN (0.5 mol/L in THF, 2.5 mL, 1.25 mmol), 3 mol/L of aq NaOH (1 mL), and 30% aq H₂O₂ (1 mL) according to the procedure described for the preparation of 13 to give (3*S*,4*S*,5*R*)-*N*-tert-butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4-hydroxy-5-(hydroxyethyl)piperidine (113 mg, 91%); $[\alpha]^{23}_{\rm D}$ +5.3 (*c* 1.35, CHCl₃). This compound was prepared from the diol (90 mg, 0.18 mmol) according to the procedure described for the preparation of 13 to give 16 (28 mg, 97%); $[\alpha]^{25}_{\rm D}$ -17.8 (*c* 1.13, EtOH).

(3S,4S,5R)-*N*-tert-Butoxycarbonyl-3-(tert-butyldiphenylsilyloxy)-4-hydroxy-5-methylpiperidine (ent-12). This compound was prepared from ent-9 (182 mg, 0.4 mmol), MeLi (2.4 mL, 1.04 mol/L in Et₂O, 2.5 mmol), CuCN (112 mg, 1.25 mmol), and BF₃·OEt₂ (4 mol/L in Et₂O, 2.5 mL, 1 mmol) according to the procedure described for the preparation of **12** to give *ent*-**12** (133 mg, 71%); $[\alpha]^{25}_{D}$ +17.2 (*c* 1.03, CHCl₃).

(3S,4S,5R)-3,4-Dihydroxy-5-methylpiperidine (17). This compound was prepared from *ent*-12 (127 mg, 0.27 mmol) according to the procedure described for the preparation of **3** to give 17 (34 mg, 96%); $[\alpha]^{25}_{D}$ –7.4 (*c* 0.90, EtOH).

(3S,5S)-*tert*-Butyl 3,4,5-Trihydroxypiperidine-1carboxylate (22) and (3R,4s,5S)-tert-Butyl 3,4,5-trihydroxypiperidine-1-carboxylate (23). To a solution of ent-8 (219 mg, 0.5 mmol) in acetone (2 mL) was added an aqueous 4% OsO₄ solution (65 μ L, 0.01 mmol). After 10 min, an aqueous 50% NMO solution (176 mL, 0.75 mmol) was added and the mixture was stirred overnight. To the solution were added Na₂-SO₃ and Na₂SO₄. The mixture was filtered through a Celite pad and the filtrate was evaporated. The residue was chromatographed on silica gel (*n*-hexane:AcOEt = 1:1) to give a diastereoisomeric mixture of N-tert-butoxycarbonyl-5-tert-butyldiphenylsilyloxy-3,4-dihydroxypiperidine (236 mg, 99%). To a solution of the above diol (513.8 mg, 1.09 mmol) in THF (10 mL) was added 1 M TBAF (2,2 mL, 2.2 mmol) in THF at room temperature. After being stirred overnight, sat. K₂CO₃ (5 mL) was added and the reaction mixure was extracted with CHCl₃. The aqueous layer was extracted with CHCl₃. The combined organic solvents were dried over MgSO₄ and evaporated. The residue was chromatographed on silica gel (CHCl₃:MeOH = 1:1) to give 23 (7.4 mg, 3%) and 22 (189 mg, 74%). 22: $[\alpha]^{22}$ _D +24.5 (c 1.74, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.44 (9H, s), 2.57 (1H, br s), 2.73 (1H, br s), 3.01 (1H, d, J = 12.4 Hz), 3.50 (1H, d, J = 7.1 Hz), 3.83 (1H, br s), 3.98-4.05 (3H, m),4.21-4.43 (1H, m). ¹H NMR (400 MHz, CD₃OD) δ 1.45 (9H, s), 2.97, 3.12 (1H, br s), 3.30 (2H, dd, J = 3.2, 1.7 Hz), 3.52-3.86 (4H, m). ¹³C NMR (100 MHz, CDCl₃) δ 28.4, 29.7, 47.7, 67.5, 67.9, 74.9, 80.5, 156.0. IR (KBr) (cm⁻¹) 3371 (-OH), 2966, 2932, 2876, 1682 (C=O), 1462, 1428, 1366, 1246, 1168, 1073. EI-MS (m/z) 233 (M⁺). HRMS calcd for C₁₀H₁₉NO₅ 233.1263, found 233.1242.

23: ¹H NMR (400 MHz, CDCl₃) δ 1.45 (9H, s), 3.28 (2H, d, J = 13.2 Hz), 3.74–4.28 (7H, m). ¹H NMR (400 MHz, CD₃OD) δ 1.45 (9H, s), 3.16 (1H, br s), 3.29–3.65 (5H, m), 3.85–3.86 (1H, m). ¹³C NMR (100 MHz, CDCl₃) δ 28.3, 46.0, 68.8, 70.3, 80.5, 155.9. IR (KBr) (cm⁻¹) 3416 (–OH), 2929, 1693 (C=O), 1540, 1464, 1416, 1367, 1325, 1264, 1232, 1168, 1076, 1017. EI-MS (*m/z*) 233 (M⁺). HRMS calcd for C₁₀H₁₉NO₅ 233.1263, found 233.1286.

(3S,5S)-Piperidine-3,4,5-triol (18). A solution of 22 (179.5 mg, 0.38 mmol) in dioxane (1 mL) and 10% HCl (4 mL) was refluxed for 1 h. Et₂O (10 mL) was added to the solution and the mixture was separated. The organic layer was extracted with water and the combined aqueous layers were concentrated to yield the hydrochloride of 18 as a solid. The solid was purified by ion-exchange resin (Amberlite IRA-410, OH⁻) to give the free amine 18 (49.6 mg, 98%).

18-HCl: mp 198–199 °C [lit.^{25c} mp 195.5–196 °C]. [α]²⁵_D +20.8 (*c* 1.4, CH₃OH), [lit.^{25c} [α]²⁰_D +22.7 (*c* 0.8, CH₃OH)]. ¹H NMR (400 MHz, D₂O) δ 2.93 (1H, dd, J = 12.7, 8.5 Hz), 3.18 (1H, dd, J = 12.9, 1.7 Hz), 3.27 (1H, dd, J = 12.9, 5.9 Hz), 3.31 (1H, dd, J = 12.7, 3.9 Hz), 3.75 (1H, dt, J = 7.7, 1.3 Hz), 4.04–4.09 (1H, m), 4.21 (1H, ddd, J = 5.9, 2.7, 1.7 Hz). ¹³C NMR (67.8 MHz, D₂O, reference acetone δ 30.3) δ 45.5, 46.1, 64.7, 65.0, 70.8. IR (KBr) (cm⁻¹) 3343 (–OH), 3088, 2904, 2852, 1610, 1475, 1441, 1408, 1368, 1339, 1306, 1271, 1227, 1208, 1099, 1079, 1056. EI-MS (*m*/*z*) 133 (M⁺). HRMS calcd for C₅H₁₁-NO₃ 133.0739, found 133.0730.

18: $[\alpha]^{20}_{D}$ +65.8 (c 0.9, CH₃OH) [lit.²³ $[\alpha]_{D}$ +66.7 (c 0.3, CH₃-OH)]. ¹H NMR (400 MHz, D₂O) δ 2.31 (1H, dd, J = 12.9, 9.5 Hz), 2.62 (1H, dd, J = 14.1, 2.0 Hz), 2.82 (1H, dd, J = 13.9, 3.4 Hz), 2.96 (1H, dd, J = 13.7, 4.4 Hz), 3.45 (1H, dd, J = 8.5, 3.2 Hz), 3.67 (1H, dt, J = 9.0, 4.6 Hz), 3.87 (1H, m). ¹³C NMR (100 MHz, D₂O, reference acetone δ 30.3) δ 47.4, 47.5, 67.2, 67.3, 72.8. IR (neat) (cm⁻¹) 3388 (-OH), 2928, 2364, 2344, 1736, 1720, 1656, 1640, 1561, 1545, 1452, 1345, 1274, 1072.

EI-MS (m/z) 133 (M⁺). HRMS calcd for C₅H₁₁NO₃ 133.0739, found 133.0739.

(3*R*,5*R*)-Piperidine-3,4,5-triol (21). The piperidinetriol 21 was prepared from 8 according to the same procedure described for 18, via *ent*-22.

ent-22: (79%); $[\alpha]^{28}_{D}$ -25.0 (c 1.33, CHCl₃).

21-HCl: $[\alpha]^{25}_{D} - 20.7$ (c 1.1, CH₃OH) [lit. ^{25c} $[\alpha]^{20}_{D} - 22$ (c 0.8, CH₃OH)].

21: (98%); $[\alpha]^{20}_{D}$ -66.0 (*c* 1.2, CH₃OH).

(3R,4s,5S)-tert-Butyl 3,4,5-trihydroxypiperidine-1-car**boxylate** (23) and (\pm) -22. A solution of OsO₄ (254.2 mg, 1 mmol) in CH_2Cl_2 (1 mL) was added to a solution of (±)-3 (199 mg, 1 mmol) and TMEDA (116 mg, 1 mmol) in CH₂Cl₂ (22 mL) at -78 °C. The mixture was stirred at the same temperature for 2 h, warmed to room temperature, and stirred for 2 h. The reaction mixture was evaporated. The resulting residue was dissolved in MeOH (10 mL). Hydrochloric acid (conc 7 drops) was then added and the solution was stirred for 2 h at room temperature. The reaction mixture was concentrated under reduced pressure. The residue was redissolved in MeOH (15 mL) and Et_3N (202 mg, 2 mmol) and Boc_2O (262 mg, 1.2 mmol) were added to the resulting mixture. The reaction mixture was stirred at room temperature overnight and then evaporated. The residue was purified by chromatography on silica gel $(CHCl_3:MeOH = 10:1)$ to give 23 (153 mg, 66%) and (\pm) -22 (38 mg, 16%).

(3R,4s,5S)-Piperidine-3,4,5-triol (19). A solution of 23 (179.5 mg, 0.38 mmol) in dioxane (1 mL) and 10% HCl (4 mL) was refluxed for 1 h. Et₂O (10 mL) was added to the solution and the mixture was separated. The organic layer was extracted with water and the combined aqueous layers were concentrated to yield the hydrochloride of 19 as a solid. The solid was purified by ion-exchange resin (Amberlite IRA-410, OH⁻) to give the free amine 19 (42.5 mg, 84%).

19-HCl: mp 182–184 °C [lit.^{25c} mp 185–187 °C]. ¹H NMR (400 MHz, D₂O) δ 3.10–3.15 (4H, m), 3.92–3.97 (3H, m). ¹³C NMR (100 MHz, D₂O, reference acetone δ 30.3) δ 44.1, 65.5, 68.3. IR (KBr) (cm⁻¹) 3435 (–OH), 2944, 2824, 2674, 2568, 1582, 1465, 1439, 1415, 1339, 1320, 1255, 1190, 1117, 1094. EI-MS (*m/z*) 133 (M⁺). HRMS calcd for C₅H₁₁NO₃ 133.0739, found 133.0764.

19: mp 163–165 °C. ¹H NMR (400 MHz, D₂O) δ 2.57 (2H, d, J = 9.0 Hz), 2.64 (2H, dd, J = 12.7, 4.4 Hz), 3.58–3.62 (2H, m), 3.86 (1H, s). ¹³C NMR (100 MHz, D₂O, reference acetone δ 30.3) δ 44.9, 68.6, 70.6. IR (KBr) (cm⁻¹) 3390 (–OH), 3285, 2930, 1866, 1842, 1790, 1456, 1041. EI-MS (m/z) 133 (M⁺). HRMS calcd for C₅H₁₁NO₃ 133.0739, found 133.0715.

(3S,5S)-Piperidine-3,4,5-triol (20). A mixture of (\pm) -9 (436 mg, 0.96 mmol) in dioxane (6 mL), H₂O (4 mL), and concentrated H₂SO₄ (0.64 mL) was heated at 95 °C for 3 h. The reaction mixture was concentrated and the residue was purified by ion-exchange resin (Amberlite IRA-410, OH⁻) to give the free amines **20** (69.7 mg, 55%) and (\pm)-18 (26.0 mg, 20%).

20: mp 160–162 °C. ¹H NMR (400 MHz, D₂O) δ 2.23 (2H, dd, J = 12.7, 0.7 Hz), 2.93 (2H, dd, J = 12.4, 4.9 Hz), 3.12 (2H, dt, J = 16.1, 2.2 Hz), 3.29–3.36 (1H, m). ¹³C NMR (270 MHz, D₂O, reference acetone δ 30.3) δ 49.3, 71.0, 78.4. IR (KBr) (cm⁻¹) 3405 (–OH), 3308, 2976, 2909, 2860, 2690, 1654, 1457, 1365, 1337, 1286. EI-MS (m/z) 133 (M⁺). HRMS calcd for C₅H₁₁-NO₃ 133.0739, found 133.0728.

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Supporting Information Available: Copies of ¹H NMR and ¹³C NMR spectra of all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org. JO050519J